

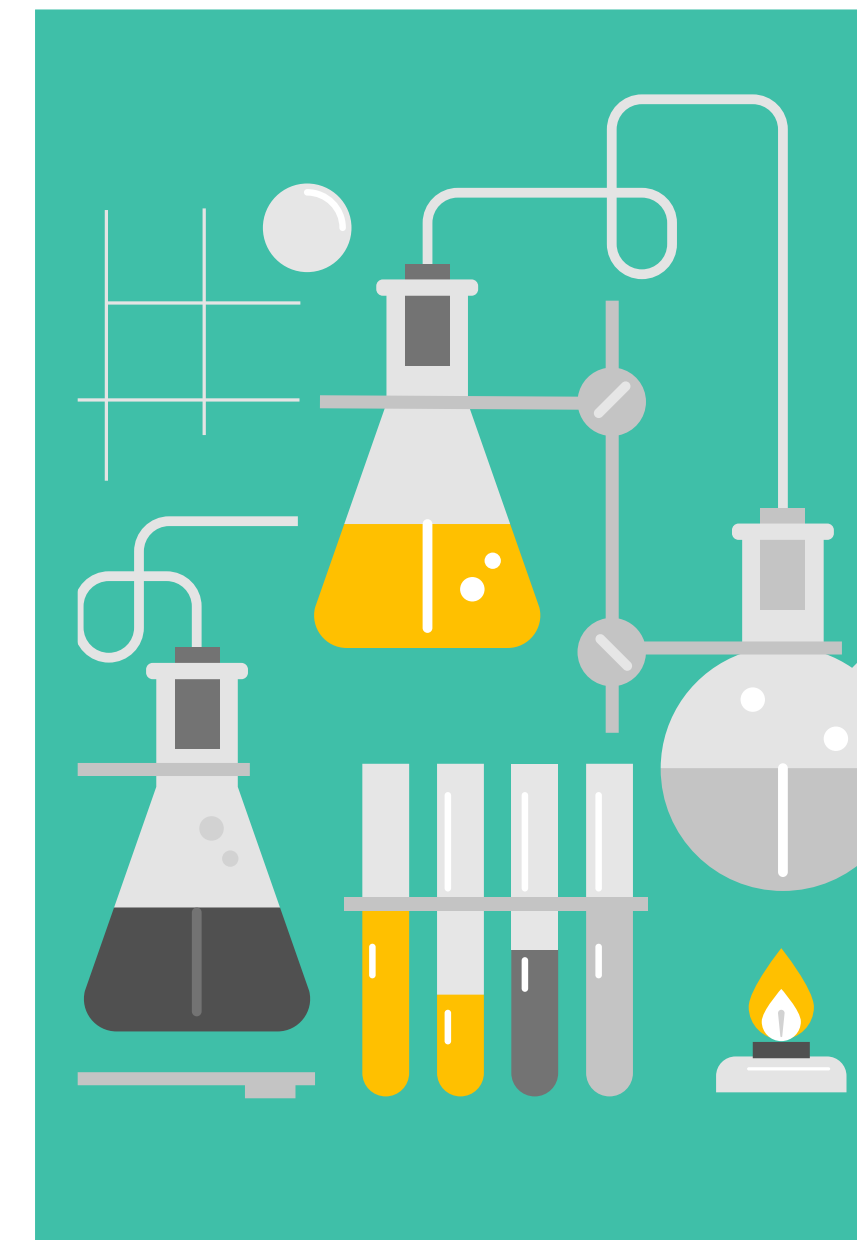
NPDN PROFICIENCY COMMITTEE

Jill Pollok¹, Chair, NEPDN; Nick Goltz², Vice-chair, NEPDN; Emran Ali³, NEPDN; John Bonkowski⁴, NCPDN; Elizabeth Bush⁵, SPDN; Ray Hammerschmidt⁶, NCPDN ; Carrie Harmon⁷, SPDN; Jeff Jones⁷, SPDN; Jennifer Olsen⁸, GPDN; Melodie Putnam⁹, WPDN; Karen Rane¹⁰, NEPDN; and Peng Tian¹¹, NCPDN

TESTIMONIALS

“Being on this committee is an opportunity to be a part of the creative process of producing assessments for diagnosticians in fun and creative ways, which also helps me gauge areas where I personally need more help. Also, I’ve been able to make relationships with experienced diagnosticians all over the country, which has proved to be one of the most valuable parts of being in a committee!”

“I’ve recently joined the proficiency committee because it deals with the improvement of plant disease diagnostic tools and techniques. The demand for advanced molecular disease diagnosis has been growing in recent years. I think being a member of this committee, I can contribute to strengthening the ongoing efforts of NPDN to increase the diagnostic capability of plant disease clinics.”



MISSION

- To provide the framework for meaningful and achievable proficiency standards and verification methods in order to ensure that NPDN diagnosticians are proficient in current diagnostic techniques and able to identify appropriate techniques.
- The proficiency standards and verification methods will improve diagnostic capability of the NPDN laboratories, while providing useful information to diagnosticians and laboratories.



- We are looking for subject matter experts to help create Proficiency Assessments
- Do you already have resources that assess diagnostic techniques or procedures? Do you want to create something new? Please submit an RFA!

High Priority Topics (we welcome topics in all subject areas though)

- Communications and Resources – procedure for samples of regulatory significance
- Conventional bacterial methods: microscopy, bacterial streaming, use of semi-selective media, pure colony isolation; Biochemical/LOPAT
- Morphological ID of fungi/ morphological ID of oomycetes
- Inputting information into the LIMS/National Data Repository
- Serological assay techniques
- Molecular analysis techniques (PCR, qPCR, RT-PCR, Sequencing, LAMP, RPA); MLSA
- Molecular analysis techniques: Sanger Sequencing, MLSA, Next-Gen Sequencing
- Plant samples: looking for feeding injury on all plant parts; frass, mites, insects, other arthropods.



2022 PLAN OF WORK

Deploy Triage module within LearnUpon:

April - committee meeting in Davis: set up “bones” of virtual Triage assessment in LearnUpon

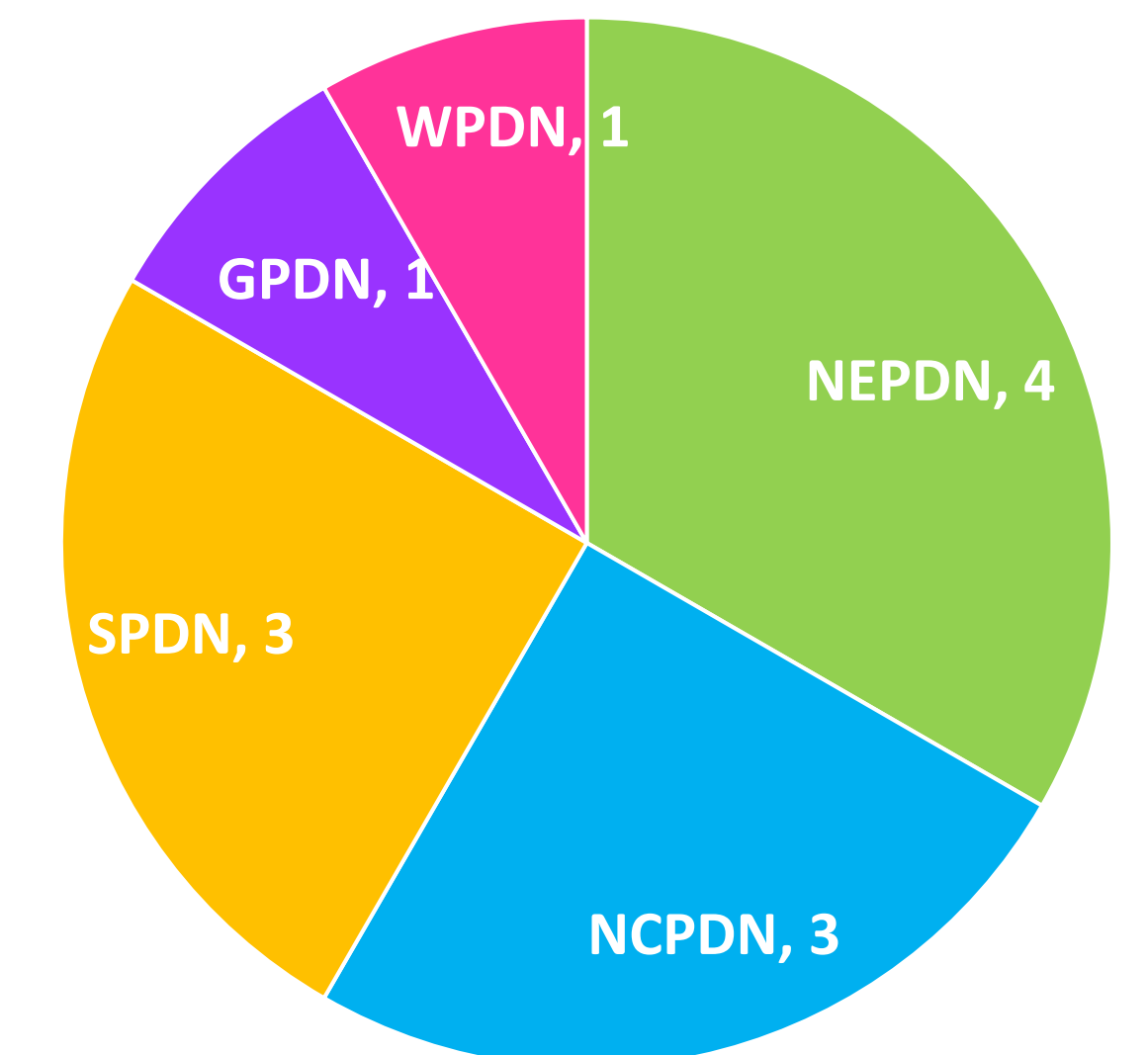
Determine assessment criteria/policy

May/Jun/Jul: Apply policy to Triage assessment and fine-tune

Fall: Continue to create assessments from high-priority list and translate them into LearnUpon



Regional Representation by number of members



- We are still looking for members from underrepresented regions: Great Plains Diagnostic Network, and Western Plant Diagnostic Network

- We welcome new ideas and creative minds from all regions!